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In silico study of dynamic level interaction between cLF36 peptide and coronavirus surface protein in Bovine coronavirus

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Abstract

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Considering the fact that coronavirus is one of the major causes of calf diarrhea syndrome, this study aimed to investigate the interaction between the peptide cLF36 and the coronavirus protein by utilizing computational methods. Coronavirus amino acid sequence was obtained and molecular docking between coronavirus and the engineered camel lactoferrin was performed. The simulation model was conducted using PROMX model. The results showed that the highest bond length was 2.5Å and the lowest 1.7Å which makes the connection between the two proteins strong, robust, and close to reality. According to the obtained results, cLF36 is a promising potential drug in the treatment of calf diarrhea.

1. Introduction

Bovine coronavirus is classified in the family Coronaviridae and specie of Betacoronavirus 1 group; with a diameter of 120-150 nm, and a positive sense single-stranded RNA, a human and animal pathogen that has always been characterized by high economic damage caused by diseases in its host [1]. This virus is the main cause of severe diarrhea in newborn calves. Additionally, it is associated with winter diarrhea in adult cows, respiratory infections in fattening cows, in addition to intestinal infections in cattle and wild ruminants which eventually causes severe losses to both dairy and fattening cattle sectors worldwide [2-5]

One of the relatively new areas of bioinformatics under the structural and modeling bioinformatics domain is the field of molecular dynamics simulation that has begun to flourish since the 70s [6]. These chemo-informatic simulations are widely used in drug design and structure optimization [7].

Lactoferrin is a glycoprotein present in leukocytes and known for its biological properties [8][9] such as its antibacterial [10], and antiviral [11] effects. Camel lactoferrin consists of 689 amino acids and 17 disulfide bridges. This protein has four glycosylation sites, one in the N lobe and three in the C lobe with a disulfide bond pattern similar to that of other lactoferrins found in human and horse; however, the predicted locations of glycosylation sites are quite different and with higher biological activity [12]. The potent chimeric peptide, which has been recently derived from camel lactoferrin (cLF36), has shown promising activity as an antibacterial [13], and anticancer agent [14]. Therefore, the aim of this study was to investigate the interaction between the peptide cLF36 and the coronavirus protein to investigate its antiviral activity by utilizing computational methods.

2. Materials and Methods

2.1. Protein data collection

S protein is an important component in virus entry and pathogenesis. Another importance of the S protein is its antigenicity. Antibodies induced by the S protein are highly neutralizing and are more stable during infection than those induced by the hemagglutinin-esterase protein [15]. This protein contains an N-terminal signal peptide which primes the nascent polyprotein for import into the ER. In the ER, the S protein is extensively modified with N-linked glycans, which may provide protection against neutralizing antibodies [16].

Bovine coronavirus amino acid sequence registered at the NCBI database (Accession number AAZ95502.1) was obtained in FASTA format. Then, coronavirus spike glycoprotein(S) amino acids were acquired from the NCBI. The same tool was then used to search against the PDB database and a suitable Bovine coronavirus pattern with a similarity of 94%, and overlapping of 95% was obtained under the PDB id (4H14_A). This protein was used as a template structure for 3D modeling of bovine coronavirus.

2.2. Protein 3D structure modeling

4H14_A sequence was modeled using SWISS-MODEL [17] and downloaded in PDB format. The YASARA (Yet Another Scientific Artificial Reality Application) [18] was used to optimize the energy structure of the model. Additionally, SAVES [19] was used to verify protein structure, and the best-predicted structures were selected and used in the following stage.

2.3. Molecular docking and dynamics simulation

The predicted interaction between CLF36 and the selected protein model was investigated using ClusPro 2.0 [20] and the best model with the most negative center score was selected. Molecular dynamics simulation procedure was carried out using the GROMACS package, which consisted of GRO file preparation, solvent addition, electronic charge neutralization, system energy minimization, temperature and pressure control, and the md file generating.

3. Results and Discussion

SWISS-MODEL predicted 3D structure results showed that there were 27 patterns and the best structure for our protein was 4H14_A, which was downloaded and saved in FASTA format and 3D photo [21]. Energy minimization was executed on YASARA server to obtain an optimized model structure with -23619 Kj/mol of initial energy to a final energy of -30920 Kj/mol. The energy minimized model of 4H14_A protein was considered structural validation studies by various online tools and software like PROCHECK, ERRAT, and PROVE [22][23]. SAVES results showed that the model was better before the implementation of YASARA; therefore, the first model was used for the docking phase (Fig. 1).

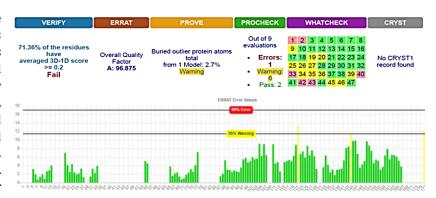


Figure 1. SAVES results for the first selected model. The results of the SAVES software were compared between the model obtained from SWISS-MODEL and the model after YASARA. Swiss model results were better in terms of overall quality, the warning percentage, and Ramachandran plot. Estimated overall quality factor by ERRAT was 96.875

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Ramachandran plot (Fig. 2) showed that 90% of the amino acids in the selected model were plotted in the most favoured region with no residues in the disallowed regions, which indicated that the chosen model was of high quality.

Z-score deviation analysis for the modeled protein demonstrated that the average statistical Z-score value was 0.535, which showed good results (Fig. 3) [19]

The overall estimated quality factor of ERRAT was 96.875 which is evocative of the fact that the structure is of good quality having high resolution with insignificant error standards of individual amino acid residues in modeled protein [25] (Fig. 1)

ClusPro 2.0 docking showed that the lowest energy level was - 854.6 which was attributed to complex (no.9), which was considered the best complex. The amount of complex energy was calculated based on the following formula [20]:

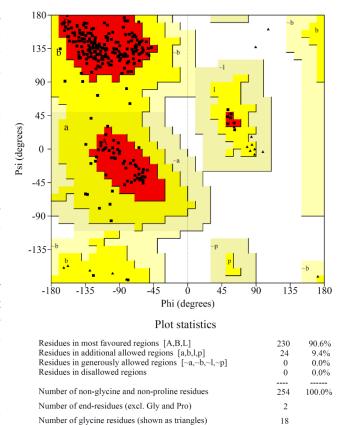
$$E = 0.4E_{rep} - 0.4E_{att} + 600E_{elec} + 1.00E_{DARS}$$

and showed that links lengths between the selected model and the peptide ranged between 1.7Å and 2.5Å (Fig. 4 A). These results indicated that there was a strong association between coronavirus peptide and cLF36 [26].

GROMACS results showed that the electrical charge was positive (10.99). Energy minimization was achieved reaching the 729 phase. Xmgrace results showed that the system was in equilibrium in terms of temperature and pressure and the final structure (Fig. 4 B) illustrated that the relationship between the two proteins was strong and robust. Therefore, this system can be suggested as a suitable model in drug design.

4. Conclusion

Bovine coronavirus is a livestock pathogen that has always been known to cause high economic losses. In this study, the association between engineered cLF36 peptide and bovine coronavirus protein was assessed computationally. The results showed that the best complex with the lowest binding energy and connection between the two proteins was strong (1.7-2.5 Å) which is close to reality. Further laboratory tests are required for final approval of cLF36 peptide in the fight against calf diarrhea.



Based on an analysis of 118 structures of resolution of at least 2.0 Angstroms and R-factor no greater than 20%, a good quality model would be expected to have over 90% in the most favoured regions.

Figure 2. The Ramachandran plot of the selected model. 90.6% of amino acids were located in the most favoured region of the chart (red) which shows that the obtained model was of high quality.

* The plot was generated using PROCHECK [24]

Number of proline residues

Total number of residues

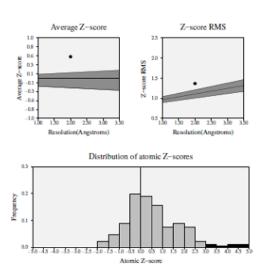


Figure 3. Z-score value calculated via PROVE software.

Z-score mean: 0.535, Z-score SD: 1.261, Z-Score RMS: 1.368, Outliers: 2.7%.

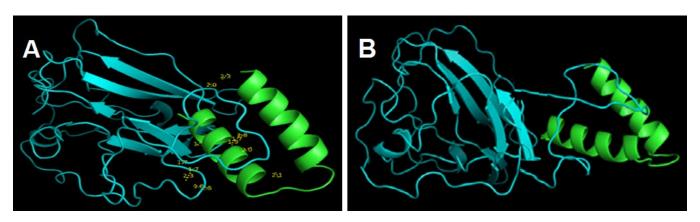


Figure 4. Docking results between the protein model (blue) and cLF36 (Green)(A) and the final structure of the system from Gromax (B). The link length ranged between 1.7Å and 2.5Å indicating a strong predicted association between the protein model and cLF36.

Conflict of interest statement

The authors declared no conflict of interest.

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Data availability statement

The authors declared that all related data are included in the article.

References

- 1. Lai MM. Coronaviridae: the viruses and their replication. In: Fields Virology. Fields, B.N., Howley, P.M., DMK (eds.). Lippincott Raven, Philadelphia, USA. 2001:1163-86.
- 2. Cho KO, Hasoksuz M, Nielsen PR, Chang KO, Lathrop S, Saif LJ. Cross-protection studies between respiratory and calf diarrhea and winter dysentery coronavirus strains in calves and RT-PCR and nested PCR for their detection. Arch. Virol. 2001;146(12):2401-19.
- 3. Clark MA. Bovine coronavirus. Br. Vet. J. 1993;149(1):51-70. DOI
- 4. Saif LJ. Coronaviruses of domestic livestock and poultry: interspecies transmission, pathogenesis, and immunity. Nidoviruses. 2007:279-98. DOI
- 5. Saif LJ, Thiel K. Viral diarrheas of man and animals. CRC press; 1989:185–252.
- 6. Hospital A, Goñi JR, Orozco M, Gelpí JL. Molecular dynamics simulations: advances and applications. Adv. Appl. Bioinform. Chem. 2015;8:37. DOI
- 7. Ranjbar MM, Ataei S, Nikbakht Brujeni G, Golabdar S. Analysis of variations, structures, and phylogenic characteristics of bovine leukocyte antigen DRB3 exon2. Arch. Razi Inst. 2017;72(3):147-57.
- 8. Kanyshkova TG, Buneva VN, Nevinsky GA. Lactoferrin and its biological functions. Biochemistry. 2001;66(1):1-7. DOI
- 9. Shahidi F, Roshanak S, Javadmanesh A, Tabatabaei Yazdi F, Pirkhezranian Z, Azghandi M. Evaluation of antimicrobial properties of bovine lactoferrin against foodborne pathogenic microorganisms in planktonic and biofilm forms (in vitro). J. Consum. Prot. Food S. 2020:1-7. DOI

- 10. Kieckens E, Rybarczyk J, Cox E, Vanrompay D. Antibacterial and immunomodulatory activities of bovine lactoferrin against *Escherichia coli* 0157: H7 infections in cattle. Biometals. 2018 Jun 1;31(3):321-30. DOI
- Ikeda M, Sugiyama K, Tanaka T, Tanaka K, Sekihara H, Shimotohno K, Kato N. Lactoferrin markedly inhibits hepatitis C virus infection in cultured human hepatocytes. Biochem. Biophys. Res. Commun. 1998;245(2):549-53.
 DOI
- Redwan EM, EL-Fakharany EM, Uversky VN, Linjawi MH. Screening the anti infectivity potentials of native N-and C-lobes derived from the camel lactoferrin against hepatitis C virus. BMC Complement Altern. Med. 2014;14(1):219. DOI
- 13. Daneshmand A, Kermanshahi H, Sekhavati MH, Javadmanesh A, Ahmadian M. Antimicrobial peptide, cLF36, affects performance and intestinal morphology, microflora, junctional proteins, and immune cells in broilers challenged with *E. coli*. Sci. Rep. 2019;9(1):1-9. DOI
- 14. Tanhaiean A, Jaafari MR, Ahmadi FS, Vakili-Ghartavol R, Sekhavati MH. Secretory expression of a chimeric peptide in *Lactococcus lactis*: assessment of its cytotoxic activity and a deep view on its interaction with cell-surface glycosaminoglycans by molecular modeling. Probiotics Antimicrob. Proteins. 2019;11(3):1034-41. DOI
- 15. Lin XQ, O'reilly KL, Storz J, Purdy CW, Loan RW. Antibody responses to respiratory coronavirus infections of cattle during shipping fever pathogenesis. Arch. Virol. 2000;145(11):2335-49. DOI
- 16. Walls AC, Tortorici MA, Frenz B, Snijder J, Li W, Rey FA, DiMaio F, Bosch BJ, Veesler D. Glycan shield and epitope masking of a coronavirus spike protein observed by cryo-electron microscopy. Nat. Struct. Biol. 2016;23(10):899. DOI
- 17. Waterhouse A, Bertoni M, Bienert S, Studer G, Tauriello G, Gumienny R, Heer FT, de Beer TA, Rempfer C, Bordoli L, Lepore R. SWISS-MODEL: homology modelling of protein structures and complexes. Nucleic Acids Res. 2018;46(W1):W296-303. DOI
- 18. Land H, Humble MS. YASARA: a tool to obtain structural guidance in biocatalytic investigations. InProtein Engineering. Humana Press, New York, NY. 2018:43-67. DOI
- 19. Pontius J, Richelle J, Wodak SJ. Deviations from standard atomic volumes as a quality measure for protein crystal structures. J. Mol. Biol. 1996;264(1):121-36.
- 20. Kozakov D, Hall DR, Xia B, Porter KA, Padhorny D, Yueh C, Beglov D, Vajda S. The ClusPro web server for protein–protein docking. Nat. Protoc. 2017;12(2):255. DOI
- 21. Kopp J, Schwede T. The SWISS-MODEL Repository of annotated three-dimensional protein structure homology models. Nucleic Acids Res. 2004;32(suppl_1):D230-4. DOI
- 22. Dave ME, Daga AD, Rawal RA. Structural and functional analysis of AF9-MLL oncogenic fusion protein using homology modeling and simulation based approach. Int. J. pharm. Pharm. sci. 2015;7(12):155-61.
- 23. e Zahra SN, Khattak NA, Mir A. Comparative modeling and docking studies of p16ink4/Cyclin D1/Rb pathway genes in lung cancer revealed functionally interactive residue of RB1 and its functional partner E2F1. Theor. Biol. Med. Model. 2013;10(1):1. DOI
- 24. Laskowski RA, MacArthur MW, Moss DS, Thornton JM. PROCHECK: a program to check the stereochemical quality of protein structures. J. Appl. Crystallogr. 1993;26(2):283-91. DOI
- 25. Colovos C, Yeates TO. Verification of protein structures: patterns of nonbonded atomic interactions. Protein Sci. 1993;2(9):1511-9. DOI
- 26. Kozakov D, Beglov D, Bohnuud T, Mottarella SE, Xia B, Hall DR, Vajda S. How good is automated protein docking?. Proteins. 2013;81(12):2159-66. DOI