

# Antibacterial activity of *Capparis spinosa* honey against *Staphylococcus aureus* and *Escherichia coli*

Abdelkrim M. Amer<sup>1\*</sup>; Saleh A. M. Abid-Alla<sup>2</sup>

1, Department of Zoology at the University of Benghazi, Benghazi, Libya

2, Plant Protection Department at Omar Al-Mukhtar University, Al Bayda, Libya

## Abstract

### E-mail:

[abdeabdelkrim.amer@uob.edu.ly](mailto:abdeabdelkrim.amer@uob.edu.ly)

Received: 15/08/2020

Accepted: 10/10/2020

Available Online: 16/10/2020

Published: 01/01/2021

**Keywords:** Caper honey, Antibacterial, *Capparis spinosa*, *Escherichia coli*, *Staphylococcus aureus*

This study aimed to investigate the antibacterial effect of caper honey collected from Jabal Al Akhdar region in eastern Libya. Honey was diluted with sterile distilled water at various rates (15%, 25%, 50%, and 75%). The dilutions were tested *in vitro* against two types of bacteria, one of which was Gram-negative (*Staphylococcus aureus*) and the other Gram-positive (*Escherichia coli*), by conducting the sensitivity tablets test using the disk diffusion method and measuring the diameter of the inhibition zone due to the effect of honey. Additionally, honey effect on the numbers of bacterial cells was investigated using the decimal dilution procedure. Caper honey analysis showed that moisture content was 18.9, with a small amount of calcium and phosphorus, and a protein content of 0.830%. Caper honey showed an apparent inhibitory effect on the studied bacteria as there was a direct relationship between honey concentration and the diameter of the inhibition zone. Additionally, there was a significant negative correlation between honey concentration and the total number of bacterial cells with total inhibition under 75% concentration.

## 1. Introduction

Honey is the substance produced by honey bees after collecting plants' nectar and sugary secretions, where it undergoes a series of enzymatic reactions before being regurgitated and stored in the hexagonal wax cells. The chemical composition of honey is complex and rich. Honey contains various sugars, especially monosaccharides such as glucose and fructose. Additionally, it contains different levels of proteins, amino acids, enzymes, organic acids, and mineral substances based on its source [1]. Due to its high nutritional value and attractive flavor, honey is considered a valuable component in the human diet [1].

Honey has the ability to fight food-borne pathogens such as *Escherichia coli*, salmonella, and other bacteria, including *Staphylococcus aureus* and *Pseudomonas aeruginosa* [2] with many reports of its antimicrobial and antifungal activities [3][4]. This activity might be attributed to its osmotic pressure; the low pH, which ranges from 3.2 to 4.5 [5]; hydrogen peroxide content; the antimicrobial peptide defensin-1, which is synthesized in the bee's salivary glands [6]; as well as the presence of phytochemical agents [3][4]. Based on its original source, honey showed antibacterial activity, especially against wound infections and inflammation [7]. With antibiotic resistance becoming a serious issue nowadays [8], honey is being rediscovered as an effective wound care product [9]. Therefore, honey is considered a convenient antimicrobial therapy due to its relatively low cost and local availability [10].

Medicinal plants are the oldest sources of pharmacologically active compounds. Although synthetic pharmaceuticals are widely produced and consumed, medicinal plants continue to meet the therapeutic needs to play a dominant role in introducing new therapeutic agents. Caper (*Capparis spinosa* L.) is known for its biologically active compounds, especially polyphenols, and it has been used for centuries in Indian traditional medicine as a medicine for various human diseases [11]. Caper honey is a dark brown color, slow-freezing honey reported to increase immunity experimentally [12].



Considering the previously mentioned points, this study aims to:

1. Investigating the chemical composition of *Capparis spinosa* honey.
2. Investigating capris honey antibacterial effect against *Staphylococcus aureus* and *Escherichia coli*.

## 2. Material and Methods

### 2.1. The Honey

Caper (*Capparis spinosa* L.) honey was sampled from the middle of May to the middle of September.

### 2.2. The Bacteria

This study concentrated on two bacterial species, *Escherichia coli* and *Staphylococcus aureus* since these bacteria are responsible for various infections such as urinary tract infection (UTI) and Post-operative wound infection. The bacterial strains were obtained from Ibin-Zahr laboratory in the form of 24-hour-old bacterial broth cultures in sterilized glass bottles with tight lids. The bacterial samples were kept at 4 °C until secondary culturing before the experiment.

### 2.3. Honey analysis

Total dissolved solids were measured using a refractometer at a temperature of 20 °C. From the refractive index readings, the percentage of water content in honey samples was estimated using the relationship between refractive index and water content [13].

The pH of honey was measured using a pH meter (Wissenenschagl-Tech, Werstaten, Wtw, Microprocessor pH meter) [13].

Determination of total sugars, vitamins, and protein content in honey samples was performed using (UV) method. For this purpose, 10 g of well-mixed honey was placed in a glass container and heated for 15 minutes at a temperature of 60 °C, with constant stirring. Honey samples were then left to cool down. 1 g the treated honey samples were weighed and placed in a volumetric flask and diluted with distilled water to 100 ml. 0.1 ml of the final solution was used to estimate the percentage of total sugars and proteins at wavelengths of 490 and 510 nm, respectively, according to [14][15] using a UV/VIS Spectrophotometer (Thermo Fisher 8735).

Calcium, potassium, and phosphorous contents were estimated in honey samples using Electrolyte Analyzer (A&T 980 / Biomedical Instrument, Switzerland).[16]

### 2.4. Growth inhibition experiment

This test was carried out by dipping discs of sterile filter paper in sequential concentrations of caper honey solution [0 (Control), 15, 25, 50, 75%] (w:v) diluted using sterile distilled water. These discs were placed using sterile forceps on the centers of Petri dishes, which were previously filled with nutrient agar and inoculated with the studied bacteria. Three replicates (dishes) were prepared for each honey concentration, and the dishes were then incubated at 37 °C. After 24 hours, the diameter of the inhibition area around each disc was measured in millimeters.

### 2.5. Bacterial count experiment

This test was carried out using the same caper honey concentrations in the growth inhibition experiment [0 (Control), 15, 25, 50, 75%] (w:v) diluted in sterile normal saline. The bacterial suspensions were prepared by scraping an individual colony from the original culture and adding it to 10 ml of sterile normal saline, from which, serial dilutions were prepared by adding 1 ml of the original suspension to 9 ml of sterile normal saline and repeat this procedure until reaching a concentration of  $10^{-5}$ . 1 ml of the final dilution was added to a test tube containing 9 ml of Honey solution with the required concentration. Three tubes were used as replications for each concentration. The tubes were incubated for 12 hours at a temperature of 37 °C. After that, 1 ml of the honey solution containing the bacterial suspension was added to a Petri dish containing the nutrient agar medium, and the dishes were incubated for 24

hours at a temperature of 37 °C. The dishes were removed from the incubator, and the growing bacterial colonies were counted using a counter. The bacterial counts were represented in colony-forming unit per ml (Cfu/ml).

## 2.6. Statistical analysis

The results were submitted to analysis of variance (ANOVA) and means were compared using Tukey's test ( $p \leq 0.05$ ) with Minitab software.

## 3. Results and discussion

### 3.1. Caper honey analysis

The results showed that caper honey total solids content was 81.1%, and therefore, water content was 18.9%. The high moisture content is due to the high moisture content of caper flowers nectar. Additionally, the harvest season also influences honey moisture content as fall and winter honey has a higher water content than that collected in the spring and summer seasons. High moisture levels can result from poor harvesting practices, or they can indicate early harvest, which may occur when honey is immature [1]. According to the current legislation, the maximum allowable moisture content in honey is 20%. Sugar content reached 74.30%; while on the other hand, calcium, potassium, and phosphorus contents were 0.07, 0.15, and 0.02 %, respectively. Protein content was 0.83%; however, these proteins include several enzymes in small quantities and significant antibacterial activities. Therefore, this modest protein content is considered highly beneficial from a medical and nutritional point of view. The recorded pH was 4.8 (Table 1).

**Table 1. Chemical analysis of caper honey samples.**

Total solids	Moisture content	Total sugars	Protein content	Calcium	Potassium	Phosphorus	pH
(%)							
81.1	18.9	74.3	0.83	0.07	0.15	0.02	4.8

### 3.2. Antibacterial effect of caper honey

The disk diffusion experiment showed that all caper honey dilution had significantly inhibited bacterial growth compared to control treatment (Fig. 1 A). Increasing caper honey concentration was accompanied by a persistent increase in growth inhibition zone diameter for both bacterial species with 28 and 24 mm in *E. coli* and *S. aureus*, respectively, using 75% honey concentration. However, this increase was significant until 50%, with no significant differences between 50% and 75% treatments in both bacterial inhibition zone diameters. Interestingly, the inhibition effect was more pronounced against *E. coli*; however, *E. coli* inhibition zone diameter was significantly higher than that of *S. aureus* only under 50% treatment.

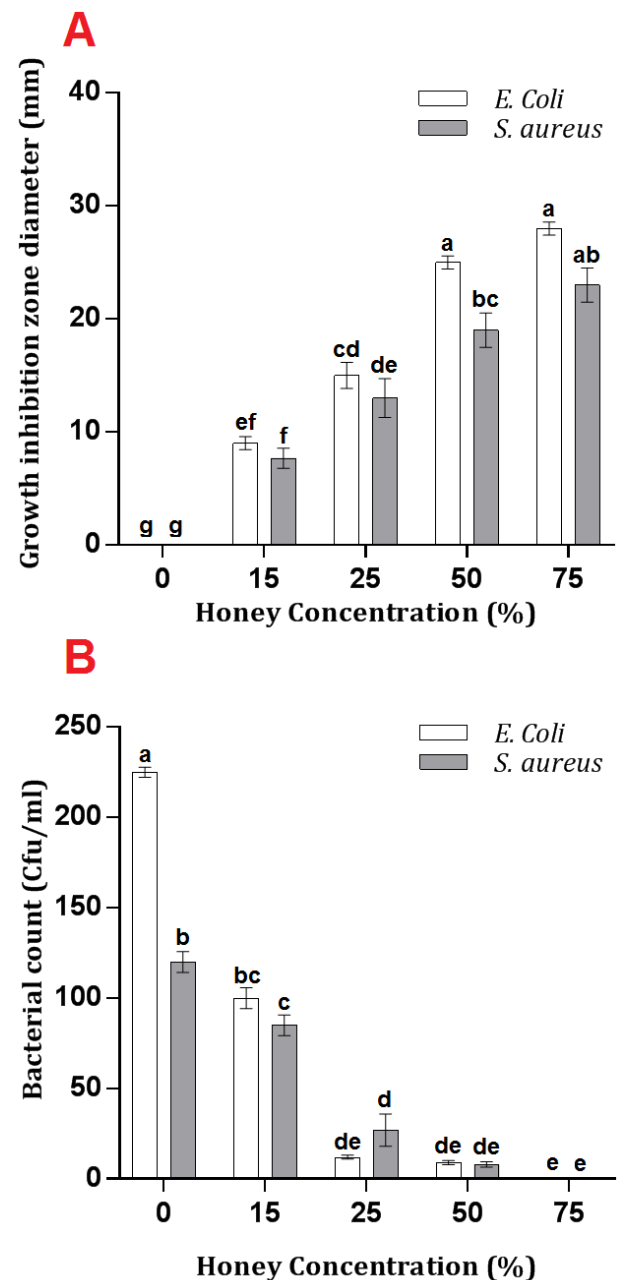
The effect of Caper honey on the numbers of study bacteria *S. aureus* and *E. coli* was investigated using plate count methods. A steady decrease in the bacterial count for both species was noticed by increasing honey concentration. The number of *E. coli* and *S. aureus* colonies decreased from 225 and 120 Cfu/ml, respectively, in control to 0 Cfu/ml using 75% caper honey concentration treatment. However, no significant decrease in *E. coli* and *S. aureus* bacterial counts after 25% and 50% treatments, respectively (Fig. 1 B).

The results of this study agreed with the results of [17][18][19], where an increased honey concentration resulted in an increase in bacterial inhibition one diameter. The results were also in correspondence with [20], who reported various inhibition degrees of *E. coli* and *Pseudomonas aeruginosa* under 5%, 10%, 20% honey concentrations. Other studies also reported the bacterial inhibitory effects of wild honey [21][22][23]. However, the current study suggested 50% and 75% caper honey concentrations as sufficient potential bacterial inhibitors under the studied conditions.

The current study nominates caper honey as an effective antibacterial agent comparable to commercial antibiotics since similar results were obtained previously by [24] using Uremia honey, and [25] using types of honey from India and New Zealand. Both studies obtained inhibition results against the growth of Gram-negative and Gram-positive bacteria similar to that of standard antibiotics.

## References

1. Richter W, Jansen C, VENZKE T, Mendonça CR, Borges CD. Avaliação da qualidade físico-química do mel produzido na cidade de Pelotas/RS Physicalchemical quality evaluation of honey produced in the municipality of Pelotas/RS. *Alim. Nutr., Araraquara.* 2012;22(4):547-53.
2. Othman AS. Antibacterial Activity of Bee and Yemeni Sidr Honey Against Some Pathogenic Bacterial Species. *Int. J. Curr. Microbiol. App. Sci.* 2014;3(10):1015-25.
3. Frankel S, Robinson GE, Berenbaum MR. Antioxidant capacity and correlated characteristics of 14 unifloral honeys. *J. Apic. Res.* 1998;37(1):27-31. DOI
4. Subrahmanyam M. A prospective randomised clinical and histological study of superficial burn wound healing with honey and silver sulfadiazine. *Burns.* 1998;24(2):157-61. DOI
5. Cooper RA, Molan PC, Harding KG. The sensitivity to honey of Gram-positive cocci of clinical significance isolated from wounds. *J. Appl. Microbiol.* 2002;93(5):857-63. DOI
6. Ilyasov R, Gaifullina L, Saltykova E, Poskryakov A, Nikolenko A. Review of the expression of antimicrobial peptide defensin in honey bees *Apis mellifera* L. *J. Apic. Res.* 2012;56(1):115-24. DOI
7. Subrahmanyam M, Hemmady A, Pawar SG. Antibacterial activity of honey on bacteria isolated from wounds. *Ann. Burn. Fire Disasters.* 2001;14(1):100.
8. Mama M, Teshome T, Detamo J. Antibacterial activity of honey against methicillin-resistant *Staphylococcus aureus*: a laboratory-based experimental study. *Int. J. Microbiol.* 2019;2019. DOI
9. Molan P. C. Using honey in wound care. *Int. J. Aromather.* 2006;3(2):21-4.
10. Zainol MI, Yusoff KM, Yusof MY. Antibacterial activity of selected Malaysian honey. *BMC Complement Altern. Med.* 2013;13(1):1-0. DOI
11. Mohamed AA, El-Beltagi HS, Ali SI. Nutraceutical Properties of *Capparis spinosa* as a Mediterranean Medicinal Plant. *Phyto-chemicals and Ethanobotany.* 2019;93(3):178-94.



**Figure 1.** The effect of *Capparis spinosa* honey on *E. coli* and *S. aureus* using disk diffusion (A) and bacterial count (B) methods.

12. Hegazi AG, Al Guthami FM, Al Gethami AF, El Fadaly HA. Beneficial effects of capparispinosa honey on the immune response of rats infected with *Toxoplasma gondii*. J Pharmacopuncture. 2017;20(2):112. DOI
13. Horwitz W. Official methods of analysis. Washington, DC: Association of Official Analytical Chemists; 1990:22-33
14. Toledo VAA, Ruvolo-Takasusuki MCC, Oliveira AJB, Chambó ED, Lopes SMS. Spectrophotometry as a tool for dosage sugars in nectar of crops pollinated by honeybees. In: Uddin J (ed) Macro to nano spectroscopy. InTech, Rijeka, 2011:269–90
15. Frasco D. Analysis of honey Color and HMF Content using a GENESYS UV-Visible Spectrophotometer. Thermo Fisher Scientific Inc. 2018.
16. Bağcı Y, Arslan D, Musa Özcan M, Dursun N. Determination of the mineral content of bee honeys produced in Middle Anatolia. Int J Food Sci Nutr. 2007;58(7):567-75. DOI
17. Adebolu TT. Effect of natural honey on local isolates of diarrhea-causing bacteria in southwestern Nigeria. Afr. J. Biotechnol. 2005;4(10):1172-4. DOI
18. Osho A, Bello O. Antimicrobial effect of honey produced by on some common human pathogens. Asian J. Exp. Biol. Sci. 2010;1:875-80.
19. Oyeleke SB, Dauda BE, Jimoh T, Musa SO. Nutritional analysis and antibacterial effect of honey on bacterial wound pathogens. J. Appl. Sci. Res. 2010;6(11):1561-5.
20. Wilkinson JM, Cavanagh HM. Antibacterial activity of 13 honeys against *Escherichia coli* and *Pseudomonas aeruginosa*. J. Med. Food. 2005;8(1):100-3. DOI
21. Basualdo C, Sgroy V, Finola MS, Marioli JM. Comparison of the antibacterial activity of honey from different provenance against bacteria usually isolated from skin wounds. Vet. Microbiol. 2007;124(3-4):375-81. DOI
22. Tamberker, DH, Rathod GN. Indian megahit honey: A Prospective antibiotic. J. Pharmacol. Toxicol. 2007;2(1):80-84.
23. Chute RK, Deogade NG, Kawale M. Antimicrobial activity of Indian honey against clinical isolates. Asiatic J Biotech Res. 2010;1:35-8.
24. Tajik H, Jalali FS, Javadi S. Comparison of antibacterial activities of natural Urmia honey and Penicillin derivatives: An *in vitro* study. J Anim Vet Adv. 2008;7(9):1097-100.
25. Devarajan, S, Venugopal S. *In vitro* activity of honey collected from India (Vellore) and New Zealand against selected clinical Pathogens. Pharmacologyonline. 2011;1:332-43.